

The growth, viability, and blood indices of broiler fed on papaya, black cumin, and mustard seed powder supplemented diets

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Abstract

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The effects of black cumin seed powder, papaya seed powder, and mustard seed powder on broiler chickens' growth and blood indices were investigated. The basal diets (starter and grower) were formulated and divided into five portions. The first portion (T1) had no supplementation; portion two (T2) had 1.1 g/100g oxytetracycline (OT) supplementation. 15g/kg papaya seed powder, mustard seed powder, and black cumin seed powder were added to portions three (T3), four (T4), and five (T5), respectively. The haematological indices were not significantly affected by the dietary treatments. The birds' relative growth rate was improved by oxytetracycline, papaya, mustard, and black cumin dietary supplementation. The serum alanine aminotransferase concentration of the broiler chickens in T1 was not statistically significantly different compared to T2, T3, and T4. Birds in treatments T1 and T2 had serum albumin concentrations that were comparable to those fed T3 and T4 supplemented diets but statistically lower ($P<0.05$) than T5. Weekly growth rate and relative growth rate were significantly improved under T3 and T4.

1. Introduction

The poultry industry's importance in terms of a country's socio-economic growth cannot be overstated, as it plays a critical role in providing animal proteins for the growing population in a timely and cost-effective manner. [1]. The use of antibiotics as growth promoters in animal production has been a source of concern due to their harmful effects on human health and their ever-increasing cost. [2]. Additionally, the possible role of antibiotic growth promoters in developing antimicrobial resistance in humans has prompted the ban of its use in many developed countries; and such activity is also recommended for other low-income and middle-income countries [2]. Furthermore, there is an increased awareness of the consumers regarding the dangers of consuming meat from animals produced with synthetic chemicals and major limits on antibiotic growth promoters in meat production in several countries [3]. Therefore, the search for alternative and safe growth-promoting compounds became widely required [4][5].

Recently, the medicinal or pharmacological properties of papaya seed, black cumin seed, and papaya seed were reported [6]. Furthermore, the effects of supplementing broiler chickens with papaya leaf and seed powders composite mix on enhancing growth efficiency characteristics and decreasing serum alanine aminotransferase and cholesterol levels were studied [7]. It was previously shown that phytogenics might affect blood parameters [8][9]. Blood haematological variables represent an effective means of assessing the health and nutritional status of animals in feeding trials [5][10]. There is currently inadequate data on the effects of supplementing broiler chickens with



papaya, mustard, and black cumin seed powders on growth, haematological indices, and serum biochemical parameters. Therefore, this study aims to investigate the effects of these phytochemicals on the aforementioned indices and parameters.

2. Materials and Methods

2.1. Collection and analysis of seeds

The papaya (*Carica papaya*) seeds were obtained from ripe fruits. The seeds were washed, drained, and shade dried. The brown mustard (*Brassica juncea*) and black cumin (*Nigella sativa*) were procured from northern Nigeria. The black cumin (BSM), papaya (PSM), and mustard (MSM) seed were milled (100µm particular size), and seed powders were obtained. The proximate compositions and phytoconstituents of prepared seed powders have been mentioned previously [11] and are shown in (Table 1).

2.2. Experimental diets

The composition of the broiler chicken basal diets formulated according to NRC specifications [12] for the starter and grower phases was stated in [11] and shown in (Table 2). After that, these basal diets were divided into five equal portions. Portion one (T1) was not supplemented; portion two (T2) was supplemented with 1.1g/100g oxytetracycline (OT). The third (T3), fourth (T4), and fifth (T5) portions were supplemented with 15g/1000g of PSM, MSM, and BSM, respectively.

2.3. Experimental procedure

All applicable National Regulations and Institutional Policies for animal care and use were followed during animal research. The Research and Ethics Committee of the Department of Animal Science, Joseph Ayo Babalola University (JABU), Ikeji Arakeji, Nigeria, granted permission to use animals and the animal protocol.

A feeding trial was performed at the JABU Teaching and Research Farm. A total of 300 Arbor Acres broiler the chicks were distributed in five treatments (10 chicks/replicate; 60 chicks/treatment) using a fully randomized experimental design. For the first seven days of brooding, the house temperature was kept at 31±2°C, but for each subsequent seven days, the temperature was lowered by 2°C until the house temperature was 26±2°C. The brooding temperature was set in this way to reduce the energy needed for temperature regulation by the chicks. During the experiment, the house was lit for 22 hours a day, and the feed and water were given *ad libitum*.

Table 1. The proximate composition and phytochemicals of mustard, papaya, and black cumin seed powders.

Parameters (g/kg)	Black cumin seed powder	Papaya seed powder	Mustard seed powder
<i>Proximate composition</i>			
Crude protein	166.81	157.84	186.82
Crude fibre	63.63	118.62	56.93
Ether extract	28.72	36.81	138.84
Ash	50.94	52.93	42.52
Nitrogen free extract	609.60	519.92	496.13
<i>Phytochemicals</i>			
Alkaloid	4.95	4.08	5.26
Flavonoids	0.06	0.08	0.07
Phenol	1.76	2.06	1.87
Saponin	1.27	3.11	1.16
Tannin	0.37	0.28	0.43

Adegbeye et al. [11].

Table 2. Composition and nutrient contents of basic diets.

Ingredients (g/kg)	Starter diet (1-21 days)	Grower diet (22-42 days)
Maize	426.60	485.60
Soybean meal	386.90	347.90
Wheat offal	121.00	101.00
Vegetable oil	22.00	23.00
Limestone	14.00	14.00
Di-calcium phosphate	18.00	17.00
Methionine	3.00	3.00
*Premix	3.00	3.00
Lysine	2.50	2.50
Salt	3.00	3.00
<i>Chemical analysis (g/kg)</i>		
Crude protein	221.30	207.80
Crude fiber	45.50	438.90
<i>Calculated analysis (g/kg)</i>		
Energy (kcal/kg)	2955.88	3000.24
Available Phosphorus	6.00	5.50
Calcium	10.20	9.30
Methionine	6.30	3.80
Lysine	11.50	10.30

Adegbeye et al. 2020 [11].

2.4. The rate of growth

A responsive weighing scale was used to assess the broiler chickens' weights at the start of the experiment (day 1) and the end of the experiment (day 42). The relative growth rate (RGR) was estimated using the following formula [13]:

$$\text{RGR} = [(w_2 - w_1) / \frac{(w_1 + w_2)}{2}] * 100.$$

Where w_1 is the weight of the broiler chickens at the onset of the experiment and w_2 is the weight of the broiler chickens on the last day of the experiment.

The viability percentage (V%) of the birds was registered daily and determined as follows [13]:

$$V \% = \left[\frac{\text{Total number of live animals at the end}}{\text{Total number of animals at the start}} \right] * 100.$$

2.5. Collection and examination of blood samples

Three birds per replication were randomly picked, and 4ml blood samples were obtained from the brachial vein using a syringe and needle. The blood was dispensed into EDTA venojets for haematological indices determination on day 21 (3 weeks mark) of the experiment. Elarabany's method [14] was used to assess the red blood cells (RBC), packed cell volume (PCV), haemoglobin concentration (Hbc), white blood cells (WBC), granulocytes (GRA), lymphocytes (LYM), and monocytes (MON). The second round of bleeding was carried out on day 42 of the experiment (6 weeks mark). About 8 ml of blood was dispensed equally into EDTA venojets and plain venojets for haematological and serum biochemical indices determination, respectively. The haematological indices were determined in the same way as it was done on day 21. The serum proteins (total protein, albumin, and globulin), as well as serum enzymes (aspartate aminotransferase [AST] and alanine transaminase [ALT]), were determined according to [5].

2.6. Analysis of the data

The following model was used in a fully randomized system: $Y_{ij} = \mu + a_i + e_{ij}$, where Y_{ij} = any of the response variables; μ = the overall mean; a_i = effect of the i th treatment (i = diets 1, 2, 3, 4 and 5) and e_{ij} = random error due to experimentation. Using SPSS version 20, all data obtained at weeks 3 and 6 were subjected to analysis of variance (ANOVA). Duncan multiple range test of the same statistical kit was used to look at the variations between treatment means.

3. Results

3.1. Phyto-genics effect on the weight, growth, and viability of the broiler chickens

The average weekly body weight gain of the birds fed with diets T3 and T5 was significantly ($P < 0.05$) higher compared to T1 and T2 (Fig. 1 A). The relative growth rate of broilers fed with T3 and T4 was not substantially different from that of broilers fed with T2 and T5 diets but was significantly higher ($P < 0.05$) than that of T1 (Fig. 1 B). Although broiler chickens fed with T3 and T4 were numerically higher than other treatments, broiler chicken viability was not significantly affected by seed powder supplementation (Fig. 1 C).

3.2. Phyto-genics effect on the haematological indices of the broiler chickens

At 3 and 6 weeks of age, no statistically significant differences between individual treatments were observed in the PCV, RBC, Hbc, mean cell haemoglobin concentration (MCHC), mean cell volume (MCV), mean cell haemoglobin (MCH), WBC, GRA, LYM, and MON (Table 3).

Table 3. Effect of different seed powder supplements on hematological indices of broiler chickens at 3 and 6 weeks of age.

Treatments	PCV (%)	RBC (x10 ⁶ /l)	HBc (g/dl)	MCHC (g/dl)	MCV (fl)	MCH (pg/cell)	WBC (x10 ⁹ /l)	GRA (x10 ⁹ /l)	LYM (x10 ⁹ /l)	MON (x10 ⁹ /l)
Age 3 week										
T1	37.00±1.62	11.50±1.24	12.33±1.54	33.31±0.05	35.06±7.72	11.68±2.73	4.72±0.88	0.88±0.16	3.75±0.52	0.08±0.02
T2	36.50±2.25	8.10±0.68	12.16±1.42	33.24±0.06	54.84±6.76	18.28±2.86	2.77±0.43	0.52±0.13	2.19±0.63	0.04±0.01
T3	34.75±3.17	9.77±0.59	11.58±1.29	33.32±0.04	37.41±8.12	12.47±2.09	2.90±0.35	0.56±0.21	2.23±0.68	0.10±0.03
T4	37.25±3.24	11.07±1.30	12.41±1.62	33.27±0.08	33.94±5.34	11.31±1.98	1.95±0.37	0.46±0.18	1.44±0.41	0.04±0.01
T5	36.25±2.98	11.35±1.31	12.08±1.77	33.25±0.05	32.65±5.77	10.88±2.54	2.62±0.09	0.70±0.11	1.85±0.39	0.06±0.02
SEM	0.90	0.57	0.30	0.02	4.06	1.35	0.28	0.07	0.22	0.01
P value	1.94	1.37	1.94	0.33	1.02	1.01	4.99	1.08	6.97	2.54
Age 6 week										
T1	33.25±2.42	9.90±0.76	11.08±1.72	33.32±0.07	42.86±9.23	14.28±3.49	4.07±0.54	2.19±0.34	1.77±0.31	0.10±0.03
T2	29.75±2.35	12.40±1.44	9.91±8.64	33.24±0.05	31.98±8.42	10.66±3.86	2.22±0.46	0.85±0.37	1.29±0.23	0.07±0.02
T3	32.75±1.89	6.20±0.52	10.91±1.62	33.32±0.04	54.14±8.99	18.05±3.72	2.82±0.39	1.13±0.30	1.59±0.25	0.09±0.03
T4	32.75±2.63	15.87±1.67	10.91±2.04	33.27±0.06	22.62±7.83	7.54±3.95	2.05±0.62	0.95±0.29	0.96±0.28	0.13±0.03
T5	33.75±2.87	13.22±1.72	11.25±1.38	33.26±0.04	33.67±8.41	11.22±3.69	2.75±0.74	0.91±0.22	1.75±0.31	0.07±0.02
SEM	0.71	1.27	0.23	0.02	4.51	1.50	0.21	0.13	0.12	0.01
P value	0.93	1.97	0.93	0.33	1.57	1.57	6.92	10.98	1.76	0.74

SEM: Standard error of the mean; PCV: Packed cell volume; RBC: Red blood cell; HBc: Haemoglobin concentration; MCV: Mean cell volume; MCH: Mean cell haemoglobin; WBC: White blood cells; GRA: Granulocytes; LYM: Lymphocytes; MON: Monocytes; SEM: Standard Error of the Mean. T1: Control; T2: Oxytetracycline (0.11g/kg); T3: Papaya seed powder (15g/kg); T4: Mustard seed powder (15g/kg); T5: Black cumin seed powder (15g/kg).

3.3. Phytochemicals effect on the serum biochemical indices of the broiler chickens

Dietary supplements had no significant effect on AST, creatinine, total protein, or globulin. The ALT serum concentration in birds fed with the diet supplemented with black cumin seed powder (T5) was statistically similar to those fed with T2, T3, and T4, but was significantly higher ($P<0.05$) than that of control (T1). The concentration of serum albumin recorded in the birds fed with T5 diet was similar to T3 and T4 but statistically higher ($P<0.05$) than T1 and T2 (Table 4).

4. Discussion

The chemical composition profiles of the phytochemicals used in this study suggest the health-supportive properties of these supplements when being used in chick's dietary programs. Secondary metabolites such as phenol and flavonoids present in the supplements have improved animal antioxidant status and antimicrobial activity [15]. Saponins have also been shown to reduce cholesterol absorption through a physicochemical interaction within the lumen [16][17]. The stability of broilers viability percentage of the birds across the various dietary treatments indicates, to some extent, the suitability of the dietary supplements and the supplementation levels used in the study for typical growth performance of the experimental birds [7][17]. The increased average weekly weight gain and relative growth rate observed in treatments T3 and T4 were consistent with previous findings [5][6]. The improved relative growth rates observed in treatments T2, T3, T4, and T5 indicate that certain phytochemicals present in the used phytochemicals have antibiotic properties comparable to oxytetracycline, resulting in reduced enteric disorder, improved health status, and improved nutrient utilization [3][7][11].

The haematological indices are reported as an indicator of normal physiological functions in animals and a reliable tool in the diagnosis of many diseases [10][18]. Considering the use of feed supplements, the obtained stability of haematological indices, including RBC, WBC, PCV, HbC, LYM, MCHC, MCV, MCH, GRA, and MON means that feed supplements did not have a detrimental impact on the usual hemopoiesis in the birds or on their normal health status at 3 and 6 weeks marks. According to [10], the WBC and their differentials (lymphocytes, granulocytes, and monocytes) are involved in fighting off infection, defending the body against foreign organisms' invasion, and in the production and transportation/distribution of antibodies. The occurrence of leukocytosis is indicative of problems such as infection, stress, trauma, allergy, or specific diseases [19]. Furthermore, it is suggested that the bacterium found in the liver, spleen, kidneys, thymus, and heart could induce the development and release of leukocytes into the bloodstream [20]. However, the steady counts of WBC recorded in broiler chickens in T1, T2, T3, T4, and T5 implies the absence of chronic infections that could have caused deviation or alteration in the normal count of the white blood cells [19].

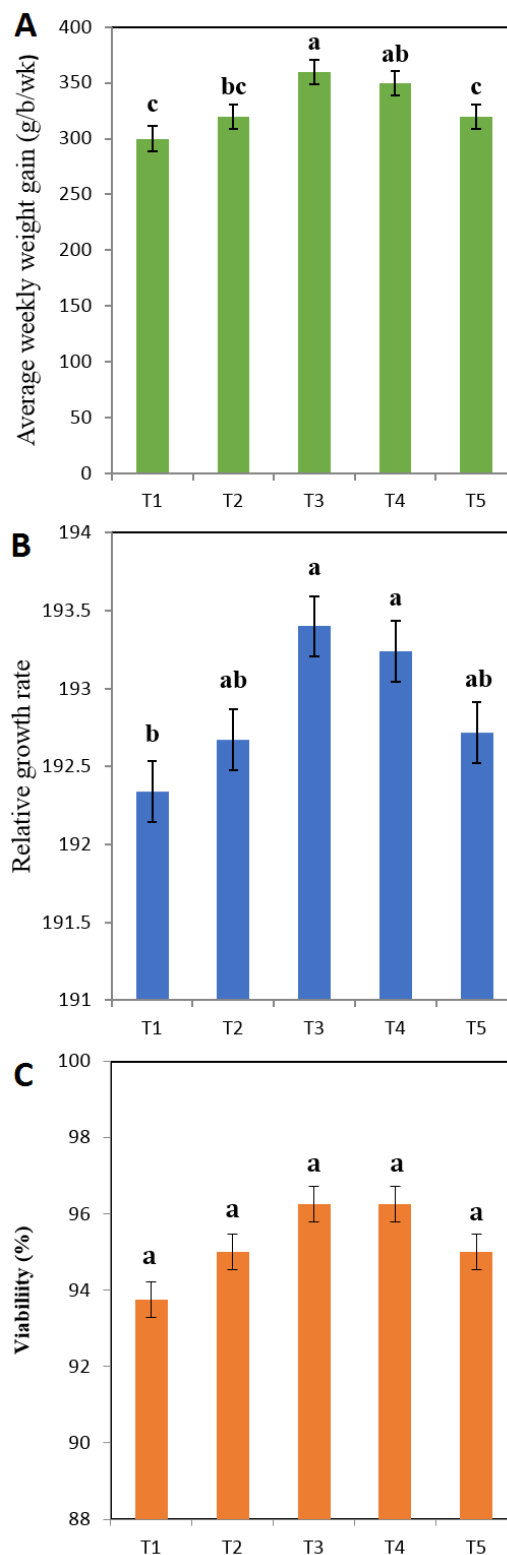


Figure 1. Average weekly weight gain (A), relative growth rate (B), and viability of broiler chickens fed with seed supplemented diets. T1: Control; T2: Oxytetracycline (0.11g/kg); T3: Papaya seed meal (15g/kg); T4: Mustard seed meal (15g/kg); T5: Black cumin seed meal (15g/kg).

Table 4. Effects of different seed powder supplements on the serum biochemical parameters at six weeks.

Treatments	AST (IU/L)	ALT (IU/L)	Creatinine ($\mu\text{mol/L}$)	Total Protein (g/L)	Albumin (g/L)	Globulin (g/L)
T1	130.75 \pm 12.23	43.05 \pm 5.67 ^b	32.45 \pm 7.39	43.63 \pm 9.85	18.60 \pm 4.82 ^b	25.03 \pm 7.54
T2	109.75 \pm 14.25	48.63 \pm 5.98 ^{ab}	40.85 \pm 6.78	39.60 \pm 8.47	20.00 \pm 5.07 ^b	19.60 \pm 6.21
T3	125.50 \pm 13.84	56.53 \pm 6.34 ^{ab}	39.25 \pm 6.33	42.78 \pm 9.08	23.40 \pm 4.33 ^{ab}	19.38 \pm 6.44
T4	144.25 \pm 16.36	57.13 \pm 6.59 ^{ab}	30.75 \pm 6.05	45.85 \pm 9.66	23.93 \pm 4.92 ^{ab}	21.93 \pm 7.22
T5	129.25 \pm 13.04	65.38 \pm 8.27 ^a	26.48 \pm 5.63	59.95 \pm 8.64	30.03 \pm 6.88 ^a	29.93 \pm 8.11
SEM	8.32	2.55	3.08	3.55	1.42	2.62
P value	0.81	0.03	0.59	0.44	0.04	0.73

Means within a column with different letters are significantly different ($P < 0.05$); AST: Aspartate Aminotransferase; ALT: Alanine Aminotransferase; T1: Control; T2: Oxytetracycline (0.11g/kg); T3: Papaya seed powder (15g/kg); T4: Mustard seed powder (15g/kg); T5: Black cumin seed powder (15g/kg).

An earlier report [21] indicated oxidative stress as the possible cause of leukocytosis, which has been observed in this study. Phytogetic supplements had been reported as a possible viable solution to the negative effect of oxidative stress in broiler chickens [7]. The potentials of phytogetic supplements in the removal or reduction of oxidative stress may explain the similar and normal WBC counts recorded in broiler chickens in this study.

The effect of age on haematological indices was reported by different authors [22][23][24]. However, the stability of the haematological indices in the current study after six weeks compared to 3 weeks mark indicated that the normal hematopoietic process in the birds was not affected by age differences.

The concentrations of ALT and AST (enzymes associated with cellular metabolism) varied between treatments. Thus, the elevated concentration of ALT in the plasma implies hepatic cell loss of integrity, indicating cellular damage [25]. The relatively higher ALT concentration recorded in treatment T5 compared to T1 (control) implies that continuous use of this dietary supplement for broiler chicken may lead to health impairment in the form of liver cells integrity compromise. The elevated ALT concentration recorded in the experimental birds fed with T5 compared to T1 may be due to the variation in the chemical composition. For instance, the flavonoids and phenol contents are relatively lower in BSM compared to PSM and MSM. The roles of flavonoids and phenol on hepatoprotective and antioxidant effects were reported in [26] and [27]. The flavonoids are highly effective polyphenolic compounds in scavenging for free radicals due to their molecule formation harboring a mobile hydrogen-containing aromatic ring with hydroxyl groups [28]. On the other hand, AST values in this study fall within the normal range (175.75-182.56 U/L) [29]. The higher serum albumin concentration recorded in birds fed with T5 suggests that BSM supplementation triggers haemo-concentration (due to dehydration or shock) and lipaemia [19]. Nonetheless, the serum albumin concentrations for broiler chickens fed with seed powder supplemented diets in this study are within the usual range (21.00–34.50 g/l) [30]. However, additional studies are required to point out the possible bioactive compounds responsible for these observations.

5. Conclusion

The various seed (papaya, mustard, and black cumin) meals have phytochemicals of pharmacological importance. The inclusion of BSM at the level 15 g/kg in the broiler chicken diet may negatively affect the liver of the birds after a relatively long period. On the other hand, weekly growth rate and relative growth rate were improved by supplementing starter and finisher phase diets with papaya and mustard seed powder at 15 g/kg in both phases.

Conflict of interest statement

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Data availability statement

The authors declared that all related data are included in the article.

References

1. Zanu HK, Asiedu P, Tampuori M, Abada M, Asante I. Possibilities of using Moringa (*Moringa oleifera*) leaf meal as a partial substitute for fishmeal in broiler chickens diets. *Online j. anim. feed res.* 2012;2(1):70-5.
2. World Health Organization. WHO guidelines on use of medically important antimicrobials in food-producing animals. 2017.
3. Valenzuela-Grijalva NV, Pinelli-Saavedra A, Muhlia-Almazan A, Domínguez-Díaz D, González-Ríos H. Dietary inclusion effects of phytochemicals as growth promoters in animal production. *J Anim Sci Tech.* 2017;59(1):1-7. [DOI](#)
4. Oloruntola OD, Ayodele SO, Oloruntola DA. Effect of pawpaw (*Carica papaya*) leaf meal and dietary enzymes on broiler performance, digestibility, carcass and blood composition. *Rev Elev Med Vet Pays Trop.* 2018;71(3):121-9. [DOI](#)
5. Oloruntola OD, Agbede JO, Ayodele SO, Oloruntola DA. Neem, pawpaw and bamboo leaf meal dietary supplementation in broiler chickens: Effect on performance and health status. *J Food Biochem.* 2019;43(2):e12723. [DOI](#)
6. Adegbeye MJ, Elghandour MM, Faniyi TO, Perez NR, Barbabosa-Pilego A, Zaragoza-Bastida A, Salem AZ. Antimicrobial and antihelminthic impacts of black cumin, pawpaw and mustard seeds in livestock production and health. *Agrofor. Syst.* 2018;94:1255–68. [DOI](#)
7. Oloruntola OD, Ayodele SO, Adeyeye SA, Jimoh AO, Oloruntola DA, Omoniyi IS. Pawpaw leaf and seed meals composite mix dietary supplementation: effects on broiler chicken's performance, caecum microflora and blood analysis. *Agrofor. Syst.* 2019;94:555–64. [DOI](#)
8. Raghuvansi SK, Prasad R, Mishra AS, Chaturvedi OH, Tripathi MK, Misra AK, Saraswat BL, Jakhmola RC. Effect of inclusion of tree leaves in feed on nutrient utilization and rumen fermentation in sheep. *Bioresour. Technol.* 2007;98(3):511-7. [DOI](#)
9. Joy PP, Thomas J, Mathew S, Skaria BP. Medicinal plants, In: Bose T K, Kabir J, Das P, Joy P P, (ed.) *Tropical Horticulture*, Naya Prokash, Calcutta, India. 2001; 449–632
10. Oloruntola OD, Ayodele SO, Agbede JO, Oloruntola DA. Effect of feeding broiler chickens with diets containing *Alchornea cordifolia* leaf meal and enzyme supplementation. *Arch. de Zootec.* 2016;65(252):489-98.
11. Adegbeye MJ, Oloruntola OD, Asaniyan EK., Agunbiade B, Oisagah EA, Ayodele SO. Pawpaw, black cumin, and mustard seed powder dietary supplementation in broiler chickens: Effect on performance, gut microflora, and gut morphology. *J Agric Sci Techn.* 2020; 22(5): 1235-46.
12. National Research Council. *Nutrient Requirement of Poultry.* 9th revised edition National Academic Press. Washington, DC, USA. 1994.

13. Adebayo FB, Adu OA, Chineke CA, Oloruntola OD, Omoleye OS, Adeyeye SA, Ayodele SO. The performance and haematological indices of broiler chickens fed Chromium Picolinate and vitamin C supplemented diets. *Asian J Res Anim Vet Sci* 2020; 6(4): 54-61.
14. Elarabany N. A comparative study of some haematological and biochemical parameters between two species from the Anatidae family within migration season. *JoBAZ*. 2018;79(31). [DOI](#)
15. Sultanbawa Y. Plant antimicrobials in food applications: Minireview. *Science against microbial pathogens: Communicating current research and technological advances*. 2011;2:1084-99.
16. Tadele Y. Important anti-nutritional substances and inherent toxicants of feeds. *Food Sci Qual Manag*. 2015;36:40-7.
17. Oloruntola OD, Ayodele SO. Pawpaw leaf meal and exo-enzyme in rabbit diet: Effect on hematological and serum biochemical indices. *Asian J. Adv. Agric. Res*. 2017;2(4):1-8 [DOI](#)
18. Togun VA, Oseni BS. Effect of low level inclusion of biscuit dust in broiler finisher diet on pre-pubertal growth and some haematological parameters of unsexed broilers. *Res. Comm. Anim. Sci*. 2005;25(1):22-4.
19. Lording PM, Friend SCE. Interpretation of laboratory results. *Aust. Vet. Pract*. 1991; 21:188-90.
20. Wati T, Ghosh TK, Syed B, Haldar S. Comparative efficacy of a phyto-genic feed additive and an antibiotic growth promoter on production performance, caecal microbial population and humoral immune response of broiler chickens inoculated with enteric pathogens. *Anim. Nutr*. 2015;1(3):213-9. [DOI](#)
21. Altan ÖZ, Pabuçcuoğlu A, Altan A, Konyalıoğlu S, Bayraktar H. Effect of heat stress on oxidative stress, lipid peroxidation and some stress parameters in broilers. *Br. Poult. Sci*. 2003;44(4):545-50. [DOI](#)
22. Hoffmann JJ, Nabbe KC, van den Broek NM. Effect of age and gender on reference intervals of red blood cell distribution width (RDW) and mean red cell volume (MCV). *Clin. Chem. Lab. Med*. 2015;53(12). [DOI](#)
23. Zakari FO, Ayo JO, Rekwot PI, Kawu MU. Effect of age, sex, physical activity and meteorological factors on haematological parameters of donkeys (*Equus asinus*). *Comp Clin Path*. 2016;25(6):1265-72. [DOI](#)
24. Rasheed AS, Olusegun OS. Influence of age and strain on hematological and blood chemistry indices on broiler chickens reared in derived savanna environment of Nigeria. *Am J Biol Life Sci*. 2017; 5:34-8.
25. Jensen JE, Stainberg SE, Freese P, Marino E. Liver function tests. *J Digest Disord*. 2004;6:1-3.
26. Xiang C, Teng Y, Yao C, Li X, Cao M, Li X, Pan G, Lu K, Galons H, Yu P. Antioxidant properties of flavonoid derivatives and their hepatoprotective effects on CCl₄ induced acute liver injury in mice. *RSC Adv*. 2018;8(28):15366-71. [DOI](#)
27. Shehab NG, Abu-Gharbieh E, Bayoumi FA. Impact of phenolic composition on hepatoprotective and antioxidant effects of four desert medicinal plants. *BMC Complement Altern Med*. 2015;15(401) [DOI](#)
28. Pannala AS, Chan TS, O'Brien PJ, Rice-Evans CA. Flavonoid B-ring chemistry and antioxidant activity: fast reaction kinetics. *Biochem. Biophys. Res. Commun*. 2001;282(5):1161-8. [DOI](#)
29. Chand N, Naz S, Rehman Z, Khan RU. Blood biochemical profile of four fast-growing broiler strains under high ambient temperature. *Appl. Biol. Chem*. 2018;61(3):273-9. [DOI](#)
30. Mitruka BM, Rawnsley HM. *Clinical biochemical and hematological reference values in normal experimental animals*. Masson Publishing. USA. 1977:142.