Effect of ammonium content in nutrition solution on vitamin C, phenols, and antioxidant capacity of three apple cultivars during cold storage

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Abstract

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ПУЗПИА

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Keywords: *Malus domestica*, Ammonium nitrate, Phenolic, Antioxidant, postharvest Pre-harvest nutritional condition is one of the major fruit quality influencers in apple whether directly after harvest or throughout cold storage. Here, the influences of various levels of ammonium to nitrate on vitamin C content, total phenolic compounds, and total antioxidant capacity of three apple cultivars ('Granny Smith', 'Gala', and 'Golab Kohans') throughout cold storage period were studied. Five different ammonium nitrogen levels (0, 0.08, 0.16, 0.19, and 0.27 meg/L) were applied. After harvest, the fruits were transferred to cold storage and stored at 0±1°C for 45, 90, and 160 days for 'Golab Kohans', 'Gala', and 'Granny Smith', respectively. Vitamin C content, total phenolic compounds, and total antioxidant capacity were determined throughout the storage period. The results showed that all cultivars showed different responses toward ammonium nitrogen application in terms of vitamin C content. 'Golab Kohans' recorded an increase in vitamin C content by increasing ammonium in nutrition solutions. On the other hand, minor changes were recorded in 'Granny Smith' and 'Gala'. Total phenolic compounds and total antioxidant capacity were significantly increased under all tested levels of ammonium in comparison to control in all cultivars. Furthermore, all cultivars showed a significant decrease in vitamin C, total phenolic compounds, and total antioxidant capacity throughout cold storage, indicating that these bioactive compounds were subjected to decomposition and metabolic degradation during the storage period. Surface response analysis results showed that more vitamin C degradation was attributed to the increase in ammonium application, while less total phenolic compounds degradation and total antioxidant capacity decrease were observed under similar treatments.

Abbreviations:

Vitamin C (VC), Total Phenolic compounds (TPC), Total Antioxidant Capacity (TAC)

1. Introduction

Apple (*Malus domestica*) is among the most popular, widely consumed, and cultivated fruit trees worldwide [1][2]. Apple fruits are usually consumed fresh or used in various food items such as desserts, jams, and drinks because of their delicious taste; in addition to the remarkable attributed health benefits [3]. Many apple cultivars are known for their ability to be stored for long periods. Due to the recent cold storage techniques, fruit markets are being supplied by apples all year round. However, fruit quality declines during cold storage and reaches minimum levels by the end of the storage period; especially those compounds of high nutritional values such as antioxidants and vitamins [4][5]. Taking into consideration the popular orientation toward healthier diets and wholesome foods [6], it is highly important to investigate the factors affecting fruit quality after harvest.

In apple nutritional programs, Nitrogen levels and source type could have substantial effects on postharvest fruit quality [7]. The unbalanced nutrition or inadequate nitrogen levels could inhibit cell division and decrease vegetative growth, which reduces the ability of plants to produce adequate yield and fruit of high quality. Therefore, choosing the proper rate of N fertilizer is crucial in apple production cycles [8][9].

Vitamin C (VC) is one of the highly important vitamins in the human diet because of its therapeutic and antioxidant properties[10] as it fortifies the body defense system against free radicals and reactive oxygen species, thereby preventing tissue damage [11][12]. Apples are considered a good source of VC as they contain considerable amounts of this vitamin [13]. Additionally, apples have elevated phenolic compounds content [14][15], which contribute significantly to the sensory characteristics such as color, astringent, and taste of apple cider and juice. Moreover, phenolic compounds have previously demonstrated vigorous antioxidant properties in several systems which refers to its cytoprotective properties [16][17]. Furthermore, apples have one of the highest levels of antioxidant activity among fruit and vegetables [18].

N fertilization has been reported to affect phenolic content in several fruits and vegetables such as Chinese cabbage [19]broccoli [20], and apple [21]. In the mentioned plants, an increase in phenolic content has been observed by reducing Nitrogen application rates. Vitamin C content and antioxidative activities in Chinese cabbage were significantly decreased with the increase in nitrogen application rates [22]. In another study, VC content was decreased by increasing the nitrogen rate in Yellow Grape Tomato nutritional program [23].

This study aimed to investigate the effect of different levels of ammonium in the nutrition solution on vitamin C, total phenolic compounds and total antioxidant capacity after harvest and during the cold storage period of three apple cultivars: 'Granny Smith' for its known storability, 'Gala', and 'Golab Kohans' for being widely cultivated in Iran as a short season fruit with good marketability but low storability potentials.

2. Material and Methods

2.1. Plant material and nutrition program

The study was carried out in 2017 and 2018 seasons in the field of the Horticultural Sciences Department, University of Tehran. The daily mean temperatures are 3° C (average low of -1° C and an average high of 8° C) in winter and 25° C (average low of 10° C and an average high of 34° C) in summer. The average humidity is 65% and 35% in winter and summer, respectively. Six years old apple trees of the cultivars 'Golab Kohans', 'Gala', and 'Granny Smith' grafted on M9 rootstock and growing in 20-liter pots under open field conditions were used in this study. Five nutrition solutions (S1, S2, S3, S4, and S5) of different ammonium to total nitrogen [NH4+/(NH4++NO3-)] were used. The ratios of ammonium nitrogen to total nitrogen were 0, 0.08, 0.16, 0.19, and 0.27 meq/L, respectively. Trees were irrigated three times in spring with irrigation solution, while in summer; they were irrigated once a week. Fruits were harvested at commercial maturity on June 29th, July 21st, and September 18th for GK, G, and GS, respectively, and transferred to cold storage at $0 \pm 1^{\circ}$ C with 90–95% relative humidity. Further information regarding agricultural practices, nutrition solutions... etc. were described in [7]

2.2. Biochemical evaluation

Vitamin C (VC), total phenolic compounds (TPC) and total antioxidant capacity (TAC) were measured throughout cold storage period at 0, 15, 30, and 45 days for GK; 0, 30, 60, and 90 days for G; and at 0, 40, 80, 120 and 160 days for 'Granny Smith' with three replications(three fruits per replication) for each evaluation date. Evaluation periods were chosen due to the different storability potentials of the three cultivars i.e. 'Golab Kohans' having the least storage ability and 'Granny Smith' having the longest storing potential.

2.2.1. Vitamin C content

VC was measured by titration with iodine and potassium iodide method. First, 16 g of potassium iodide was dissolved in 1 liter of distilled water; then,1.27 g of iodine crystals was added and the solution was stirred on a heater. VC was measured by adding 5 ml filtered apple juice to 20 ml distilled water and 2 ml of 1% starch solution. The formed solution was then titrated with potassium iodide iodine solution until a dark blue color appeared. The amount of VC was then calculated using the following formula [24]:

 $VC(mg/100g juice) = (0.88 \times V/5) \times 100$

Where V is the volume of Potassium iodide iodine consumed based on ml

 $VC (mg/100g) = VC \times [Fruit weight (g) / Fruit juice content (g)]$

2.2.2. Total phenols

Measurement of phenol content in fruits was carried out using the method of [25]. The fresh fruit sample of 0.5 g was completely crushed in 4 ml of 80% ethanol until homogenization. The homogenate was then centrifuged for 20 minutes at 9500 rounds per minute (rpm) and the clear supernatant solution was collected. In a clear tube, 8 ml double distilled water, 1 ml Sodium carbonate 7%, and 0.5 ml Folin solution was added to 0.5 ml of the clear supernatant fruit solution. After that, test tubes were put in a dark place for 90 minutes. Then, absorption at 625 nm was recorded using a spectrophotometer. The total phenols content was then calculated as gallic acid equivalents (GAE) in a Kg of fresh fruit tissue using a standard of gallic acid for absorption.

2.2.3. Total antioxidant capacity

Total antioxidant capacity was measured in the transparent extract obtained in TPC measurement. 3.4 ml of 2,2-diphenyl-1-picrylhydrazyl (DPPH) 0.1 mM were added to 1 ml of the clear methanol extract. The mixture was kept at room temperature and in the dark for one hour. TAC was measured based on the absorbance at a wavelength of 517 nm using a UV-Vis spectrophotometer in comparison to the standard curve. The results obtained were calculated and expressed as the percentage of neutralized DPPH radicals (%) [26].

2.3. Statistical analysis

The study was carried out in a completely randomized block design with five treatments and 3 replications. The experiment was repeated twice for 2017 and 2018 seasons. No significant differences were found between the studied seasons (data not shown); therefore, the average values of two seasons were used for further analysis. Data were analyzed using Duncan's multiple range test (DMRT) using GenStat statistical software 17th ed. at 5% probability level. Additionally, response surface analysis was performed between different ammonium levels and monitoring times to investigate the interaction effect between the two variants in terms of VC, TPC, and TAC.

3. Results and Discussion

3.1. Vitamin C (mg/100g)

Figure (1A) showed that the highest VC content was at harvest for all cultivars, 'Granny Smith' had the highest VC content (21.87 mg/100g) followed by 'Gala' (21.05 mg/100g) and 'Golab Kohans' (13.38 mg/100g). The differences in VC of the cultivars mostly related to their genotype [27].

A significant decrease in VC was observed in 'Golab Kohans' cultivar by increasing nitrogen of ammonium source. The maximum value of VC was found in S1 (control) with 13.37 mg/100g, while the minimum value was found in S5 which recorded 9mg/100g (Table 1). On the other hand, the only noticed a significant difference in 'Gala' was between S3 and S4 treatments with S4 being the highest in VC content. In GS, no significant differences were found among all treatments. In 'Gala', the highest vitamin C content was observed in S4 (16.04 mg/100g), while the lowest was recorded in S3 with a value of 14.72 mg/100g (Table 1). In 'Granny Smith' cultivar, the maximum value of VC was found in apples from trees fertilized with S3 (14.19 mgl100g), while minimum value was observed in S2 which recorded 13.75 mg/100g (Table 1). Various researches previously reported the effect of increased nitrogen application on VC content; however, the results of those researches were contradictory as some reported a significant increase in VC content by increasing nitrogen application, while others reported no significant differences or even decreases in VC content [28]. VC synthesis was reported to decrease under the excessive nitrogen application as a result of increased protein synthesis and decreased carbohydrate production which is crucial in vitamin C synthesis [6]. The results of the current study introduce evidence of different VC synthesis response in different apple cultivars under increased nitrogen of ammonium source application.

All cultivars showed a significant decrease in VC during the cold storage (Table 2), which corresponds with [29] who reported more than 50% drop in VC content of apple fruit after 6months of cold storage. Surface response analysis demonstrated that a more pronounced decrease in VC can be seen under high ammonium input in 'Golab Kohans' (Fig. 2 A) compared to the other cultivars that exhibited less relation between ammonium input level and VC throughout cold storage (Fig. 2 B and C). The decrease in VC content in storage is related to its nature of being susceptible to direct or enzymatic oxidation in comparison to other compounds during cold storage [30][31]. It appears that high ammonium input might contribute positively to VC degradation by participating in enzyme synthesis and/or enzymatic oxidation.

3.2. Total phenolic compounds (mg/kg)

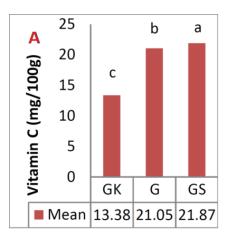
Results showed that the highest value of TPC at harvest was for 'Granny Smith' (1865 mg/kg) followed by 'Gala' (1768 mg/kg) then 'Golab Kohans' (1671 mg/kg) (Fig. 1 B).

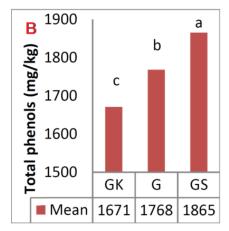
The highest values of TPC were found in S4 treatment in all cultivars with 1758, 1828, and 1942 mg/kg for GK, G, and 'Granny Smith', respectively; while the minimum values were observed in S2 for 'Golab Kohans' (1468 mg/kg) and in S1 (control) for 'Gala' and 'Granny Smith' (1582 and 1618 mg/kg, respectively) (Table 1). It was reported that 25:75% NO₃:NH₄+ treatment resulted in the highest levels of both conjugated and bound polyamines [32], which might explain the results of the current study as polyamines are key players in secondary metabolites synthesis including phenolic compounds [33]. All cultivars exhibited a significant decrease in TPC during the cold storage period (Table 2). This reduction in TPC during cold storage appears to be genotype-dependent, as it was reported that some genotypes undergo this degradation through cold storage [34][35][36]which can be interpreted as the result of phenolic compounds instability during cold storage resulting in a total metabolic degradation [35]. The surface response model illustrated that high ammonium input in all cultivars was associated with higher TPC and better preserving of the original phenolic compounds content throughout storage period compared to control (Fig. 2 D, E, and F). This effect might be attributed to ammonium water stress alleviation potentials [37] which in turn, might result in a decreased polyphenol oxidase (PPO) activity [38] and thus, fewer phenols degradation during the storage period. The studied trees might have undergone a water stress status as spring was windy and the temperature reached high values in summer at the experiment site. This might have induced water stress even with proper irrigation program. However, this needs further investigation.

3.3. Total antioxidant capacity (%)

The highest TAC at harvest was recorded in 'Granny Smith' (71.31%) followed by 'Golab Kohans' (68.04%) then 'Gala' (67.61%) (Fig. 1 C).

In regards to ammonium application, S4 gave the highest TAC for both 'Gala' and 'Granny Smith' cultivars, while S3 and S5 gave the highest TAC in 'Golab Kohans' cultivar. On the other hand, the lowest TAC values were observed under S1 treatment for all cultivars (Table 1). Similar to TPC, TAC response surface contour plotting illustrated that nutrition solutions with higher ammonium levels resulted in a better maintaining of antioxidant capacity





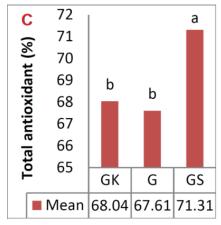


Figure 1. Effect of Cultivar on Vitamin C (mg/100g) **(A)**, total phenols (mg/kg) **(B)**, and total antioxidant (%) **(C)** in the fruits of 'Golab Kohans', 'Gala', and 'Granny Smith' apple cultivars at harvest.

Total Phenolic compounds contents were expressed as gallic acid equivalents (GAE) in a Kg of fresh fruit tissue.

Total antioxidant capacity was expressed as the percentage of neutralized DPPH radicals.

* Values were represented as an average for all treatments at harvest (regardless of ammonium input).

throughout cold storage period compared to non-ammonium nutrition solution (control) and in all cultivars (Fig. 2 G, H, and I). However, 'Golab Kohans' showed a more sharp decrease under control treatment (Fig. 2 G). The significant increase in TAC under all ammonium treatments and for all cultivars in comparison to non-ammonium control treatment (S1) might be attributed to the accumulation of polyamines due to the high ammonium concentrations [36][39]resulting in the activation of various plant defense mechanisms [40] including antioxidants [33].

All cultivars showed a significant decrease in TAC with each measurement throughout the cold storage period (Table 2) which is due to the decomposition of antioxidants throughout the storage period [41].

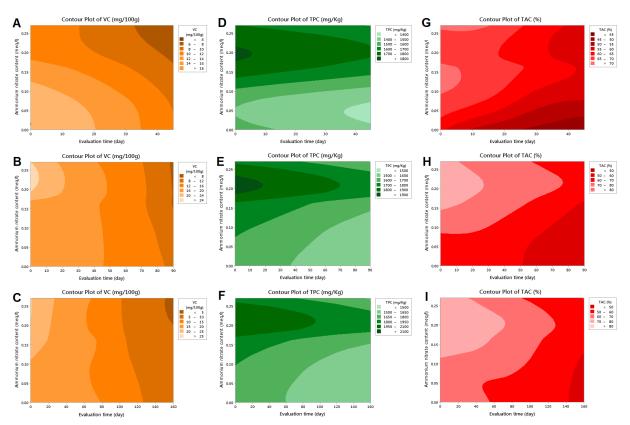


Figure 2. Surface analysis contour plotting for the relationship between ammonium content in nutrition solution and cold storage time for vitamin C content (VC) (mg/100g) (A, B, and C), total phenolic compounds (TPC) (mg/Kg) (D, E, and F), and total antioxidant capacity (TAC) (%) (G, H, and I) in the three studied apple cultivars: 'Golab Kohans' (A, D, and G), 'Gala' (B, E, and H), and 'Granny Smith' (C, F, and I).

Total phenolic compounds content were expressed as gallic acid equivalents (GAE) in a Kg of fresh fruit tissue Total antioxidant capacity was expressed as the percentage of neutralized DPPH radicals

4. Conclusion

The results of this study showed that the cultivars exhibited different responses toward ammonium nitrogen application in terms of VC content. 'Golab Kohans' recorded an increased VC content by increasing ammonium in the nutrition solution. On the other hand, minor or no changes were observed in 'Gala' and 'Grany Smith'. TPC and TACin all cultivars were significantly increased under any ammonium input in comparison to control treatment. Furthermore, all cultivars underwent a significant decrease in VC, TPC, and TAC throughout cold storage. Surface response analysis results illustrated that with the increase in ammonium inputs, more vitamin C degradation was observed. On the other hand, less TPC degradation and TAC decreasing were recorded under similar conditions.

Table 1. The effect of various levels of ammonium to total nitrogen in nutrition solutions on vitamin C content (mg/100g), total phenolic compounds (mg/kg), and total antioxidant capacity (%) of three apple cultivars ('Golab Kohans', 'Gala' and 'Granny Smith').

Cultivar	N treatment	Vitamin C (mg/100g)	total phenolic compounds (mg/kg) ^A	total antioxidant capacity% ^B
Golab Kohans	S1	13.37 ^a	1482 ^d	51.39 ^d
	S2	12.70 ^b	1468 ^d	61.72°
	S3	10.74 ^c	1710 ^b	65.44a
	S4	9.85 ^d	1758a	62.90 ^b
	S5	9.00e	1650 ^c	65.64a
Gala	S1	15.35 ^{ab}	1582 ^d	51.46 ^c
	S2	15.21 ^{ab}	1622 ^c	52.53 ^c
	S3	14.72 ^b	1708 ^b	61.24 ^b
	S4	16.03a	1828a	65.53a
	S5	15.05ab	1695 ^b	62.38b
Granny smith	S1	14.09a	1618 ^d	56.81e
	S2	13.75 ^a	1648 ^c	58.11 ^d
	S3	14.19 ^a	1774 ^b	64.98 ^b
	S4	14.04a	1942a	67.56a
	S5	14.14 ^a	1786 ^b	61.29 ^c

Different letters in each column for each cultivar indicate significant differences using Duncan multiple range test ($P \le 0.05$)

Table 2. The changes in vitamin C content (mg/100g), total phenolic compounds (mg/kg), and total antioxidant capacity(%)during cold storage period at 0 ± 1 °C and 90 - 95% relative humidity for three apple cultivars ('Golab Kohans', 'Gala' and 'Granny Smith').

Cultivar	Storage time (days)	Vitamin	total phenolic	total antioxidant
		C(mg/100g)	compounds (mg/kg) ^A	capacity% ^B
Golab Kohans	0	13.38 ^a	1671 ^a	68.04 ^a
	15	12.20 ^b	1631 ^b	63.99 ^b
	30	10.51 ^c	1594 ^c	58.95 ^c
	45	8.45 ^d	1557 ^d	54.68 ^d
Gala	0	21.05 ^a	1768a	67.61 ^a
	30	16.91 ^b	1718 ^b	62.18 ^b
	60	14.40°	1658 ^c	55.62°
	90	8.72 ^d	1603 ^d	49.10 ^d
Granny smith	0	21.87 ^a	1865 ^a	71.31 ^a
	40	17.26 ^b	1815 ^b	67.90 ^b
	80	14.07 ^c	1755 ^c	62.39 ^c
	120	8.59 ^d	1690 ^d	56.74 ^d
	160	8.42 ^d	1645 ^e	50.40e

Different letters in each column for each cultivar indicate significant differences using Duncan multiple range test ($P \le 0.05$)

A, Total phenolic compounds content were expressed as gallic acid equivalents (GAE) in a Kg of fresh fruit tissue.

B, Total antioxidant capacity was expressed as the percentage of neutralized DPPH radicals

 $A, Total\ phenolic\ compounds\ content\ were\ expressed\ as\ gallic\ acid\ equivalents\ (GAE)\ in\ a\ Kg\ of\ fresh\ fruit\ tissue$

B, Total antioxidant capacity was expressed as the percentage of neutralized DPPH radicals

Conflict of interest statement

The authors declared no conflict of interest.

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Data availability statement

The authors declared that all related data are included in the article. The nutritional compositions were previously provided as supplementary material in [7] (10.30493/das.2020.103716).

References

- 1. Wu J, Gao H, Zhao L, Liao X, Chen F, Wang Z, Hu X. Chemical compositional characterization of some apple cultivars. Food Chem. 2007;103(1):88-93. DOI
- 2. Liang W, Dondini L, De Franceschi P, Paris R, Sansavini S, Tartarini S. Genetic diversity, population structure and construction of a core collection of apple cultivars from Italian germplasm. Plant Mol. Biol. Rep. 2015;33(3):458-73. DOI
- 3. Espley R, Martens S. Health properties of apple and pear. Bioactives in fruit: health benefits and functional foods. Wiley, West Sussex. 2013:81-100.
- 4. Eberhardt MV, Lee CY, Liu RH. Antioxidant activity of fresh apples. Nature. 2000;405(6789):903-4. DOI
- 5. Suárez-Jacobo Á, Rüfer CE, Gervilla R, Guamis B, Roig-Sagués AX, Saldo J. Influence of ultra-high pressure homogenisation on antioxidant capacity, polyphenol and vitamin content of clear apple juice. Food Chem. 2011;127(2):447-54. DOI
- 6. Worthington V. Nutritional quality of organic versus conventional fruits, vegetables, and grains. J. Altern. Complement. Med. 2001;7(2):161-73. DOI
- 7. AlhajAlali F, AskariSarcheshmeh MA, Babalar M. The influence of various levels of ammonium to total nitrogen on post-harvest performance of three apple cultivars (Golab Kohans, Gala, and Granny Smith). DYSONA-Applied Science. 2020;1(1):11-9. DOI
- 8. Munson RD. Principles of plant analysis. Handbook of reference methods for plant analysis. 1998:1-24. DOI
- 9. Benitez Ramirez, M..Monitoring nitrogen levels in the cotton canopy using real-time active-illumination spectral sensing. Master's Thesis, University of Tennessee, 2010.
- 10. Okiei WO, Ogunlesi M, Azeez L, Obakachi V, Osunsanmi M, Nkenchor G. The voltammetric and titrimetric determination of ascorbic acid levels in tropical fruit samples. UNILAG Research Repository. 2009.
- 11. Xu DP, Washburn MP, Sun GP, Wells WW. Purification and characterization of a glutathione dependent dehydroascorbatereductase from human erythrocytes. Biochem. Biophys. Res. Commun. 1996;221(1):117-21. DOI
- 12. Romay CH, Armesto J, Remirez D, Gonzalez R, Ledon N, Garcia I. Antioxidant and anti-inflammatory properties of C-phycocyanin from blue-green algae. J. Inflamm. Res. 1998;47(1):36-41. DOI
- 13. Boyer J, Liu RH. Apple phytochemicals and their health benefits. Nutr. J. 2004 Dec 1;3(1):5. DOI
- 14. Khanizadeh S, Tsao R, Rekika D, Yang R, Charles MT, Rupasinghe HV. Polyphenol composition and total antioxidant capacity of selected apple genotypes for processing. J. Food Compos. Anal. 2008;21(5):396-401. DOI

- 15. Goulas V, Kourdoulas P, Makris F, Theodorou M, Fellman JK, Manganaris GA. Comparative polyphenolic antioxidant profile and quality of traditional apple cultivars as affected by cold storage. Int. J. Food Sci. 2014;49(9):2037-44. DOI
- Babbar N, Oberoi HS, Sandhu SK. Therapeutic and nutraceutical potential of bioactive compounds extracted from fruit residues. Crit. Rev. Food Sci. Nutr. 2015;55(3):319-37. DOI
- 17. Yassin LS, Alberti A, Ferreira Zielinski AA, da Rosa Oliveira-Emilio H, Nogueira A. Cytoprotective effect of phenolic extract from brazilian apple peel in insulin-producing cells. CurrNutr Food Sci. 2018;14(2):136-42. DOI
- 18. Chinnici F, Bendini A, Gaiani A, Riponi C. Radical scavenging activities of peels and pulps from cv. Golden Delicious apples as related to their phenolic composition. J. Agric. Food Chem. 2004;52(15):4684-9. DOI
- 19. Zhu W, Lin X, Jin C, Zhang Y, Fang P. Effects of nitrogen application rates on antioxidant contents and antioxidative activities in Chinese cabbage (*Brassica chinensis* L.). Zhejiang DaxueXueBaoNong Ye Yu Sheng Ming KeXue Ban. 2009;35(3):299-306.
- 20. Jones RB, Imsic M, Franz P, Hale G, Tomkins RB. High nitrogen during growth reduced glucoraphanin and flavonol content in broccoli (*Brassica oleracea* var. *italica*) heads. Aust. J. Exp. Agric. 2007;47(12):1498-505. DOI
- 21. Reay PF, Fletcher RH, Thomas VJ. Chlorophylls, carotenoids and anthocyanin concentrations in the skin of 'Gala'apples during maturation and the influence of foliar applications of nitrogen and magnesium. J. Sci. Food Agric. 1998;76(1):63-71. DOI
- 22. Zhu W, Lin X, Jin C, Zhang Y, Fang P. Effects of nitrogen application rates on antioxidant contents and antioxidative activities in Chinese cabbage (*Brassica chinensis* LZhejiang DaxueXueBaoNong Ye Yu Sheng Ming KeXue Ban. 2009;35(3):299-306.
- 23. Simonne AH, Fuzere JM, Simonne E, Hochmuth RC, Marshall MR. Effects of nitrogen rates on chemical composition of yellow grape tomato grown in a subtropical climate. J. Plant Nutr. 2007;30(6):927-35. DOI
- 24. Jalili M, R. Postharvest Physiology. Urmia University Jihad Publications, Fourth Edition, 2013. (In Persian)
- 25. Ough CS, Amerine MA. Methods for analysis of musts and wines. J. Wiley; 1988.
- 26. Tadolini B, Juliano C, Piu L, Franconi F, Cabrini L. Resveratrol inhibition of lipid peroxidation. Free Radic. Res. 2000;33(1):105-14. DOI
- 27. Lee SK, Kader AA. Preharvest and postharvest factors influencing vitamin C content of horticultural crops. Postharvest Biol. Technol. 2000;20(3):207-20.DOI
- 28. Mozafar A. Nitrogen fertilizers and the amount of vitamins in plants: a review. J. Plant Nutr. 1993;16(12):2479-506. DOI
- 29. Zubeckis E. Ascorbic acid content of fruit grown at Vineland, Ontario. Ascorbic acid content of fruit grown at Vineland, Ontario. 1962:90-6.
- 30. Veltman RH, Kho RM, Van Schaik AC, Sanders MG, Oosterhaven J. Ascorbic acid and tissue browning in pears (Pyruscommunis L. cvs Rocha and Conference) under controlled atmosphere conditions. Postharvest Biol. Technol. 2000;19(2):129-37. DOI
- 31. Sanmartin M, Pateraki I, Chatzopoulou F, Kanellis AK. Differential expression of the ascorbate oxidase multigene family during fruit development and in response to stress.Planta. 2007;225(4):873-85. DOI
- 32. Chen L, Liu QQ, Gai JY, Zhu YL, Yang LF, Wang C. Effects of nitrogen forms on the growth and polyamine contents in developing seeds of vegetable soybean. J. Plant Nutr. 2011;34(4):504-21. DOI

- 33. Mustafavi SH, Badi HN, Sękara A, Mehrafarin A, Janda T, Ghorbanpour M, Rafiee H. Polyamines and their possible mechanisms involved in plant physiological processes and elicitation of secondary metabolites. Acta Physiol. Plant. 2018;40(6):102. DOI
- 34. Wu ZS, Zhang M, Wang S. Effects of high pressure argon treatments on the quality of fresh-cut apples at cold storage. Food control. 2012;23(1):120-7. DOI
- 35. Carbone K, Giannini B, Picchi V, Scalzo RL, Cecchini F. Phenolic composition and free radical scavenging activity of different apple varieties in relation to the cultivar, tissue type and storage. Food Chem. 2011;127(2):493-500. DOI
- 36. Begić-Akagić A, Spaho N, Oručević S, Drkenda P, Kurtović M, Gaši F, Kopjar M, Piližota V. Influence of cultivar, storage time, and processing on the phenol content of cloudy apple juice. Croatian J. Food Sci. Technol. 2011;3(2):1-8.
- 37. Marino D, Moran JF. Can ammonium stress be positive for plant performance?. Front. Plant Sci. 2019;10:1103. DOI
- 38. Bower JP, Cutting JG, Wolstenholme BN. Effect of pre-and post-harvest water stress on the potential for fruit quality defects in avocado (*Perseaamericana* Mill.). S. Afr. J. Plant Soil. 1989;6(4):219-22. DOI
- 39. Claussen W, Brückner B, Krumbein A, Lenz F. Long-term response of tomato plants to changing nutrient concentration in the root environment—the role of proline as an indicator of sensory fruit quality. Plant Sci. 2006;171(3):323-31. DOI
- 40. Tassoni A, Franceschetti M, Bagni N. Polyamines and salt stress response and tolerance in Arabidopsis thaliana flowers. Plant Physiol. Biochem. 2008;46(5-6):607-13. DOI
- 41. Nowacka M, Śledź M, Wiktor A, Witrowa-Rajchert D. Changes of radical scavenging activity and polyphenols content during storage of dried apples. Int. J. Food Prop. 2014;17(6):1317-31. DOI